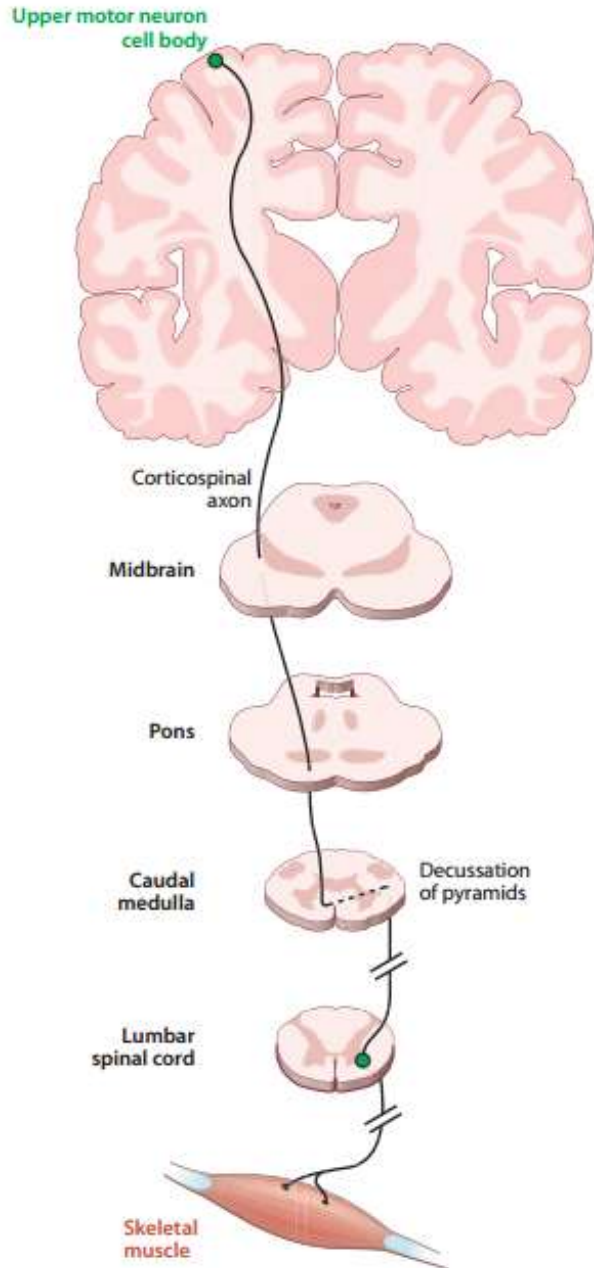


Euro HSP Annual Meeting  
17/6/2017

# Hereditary Spastic Paraplegia: **the complex form SPG9**

Emanuele Panza, PhD

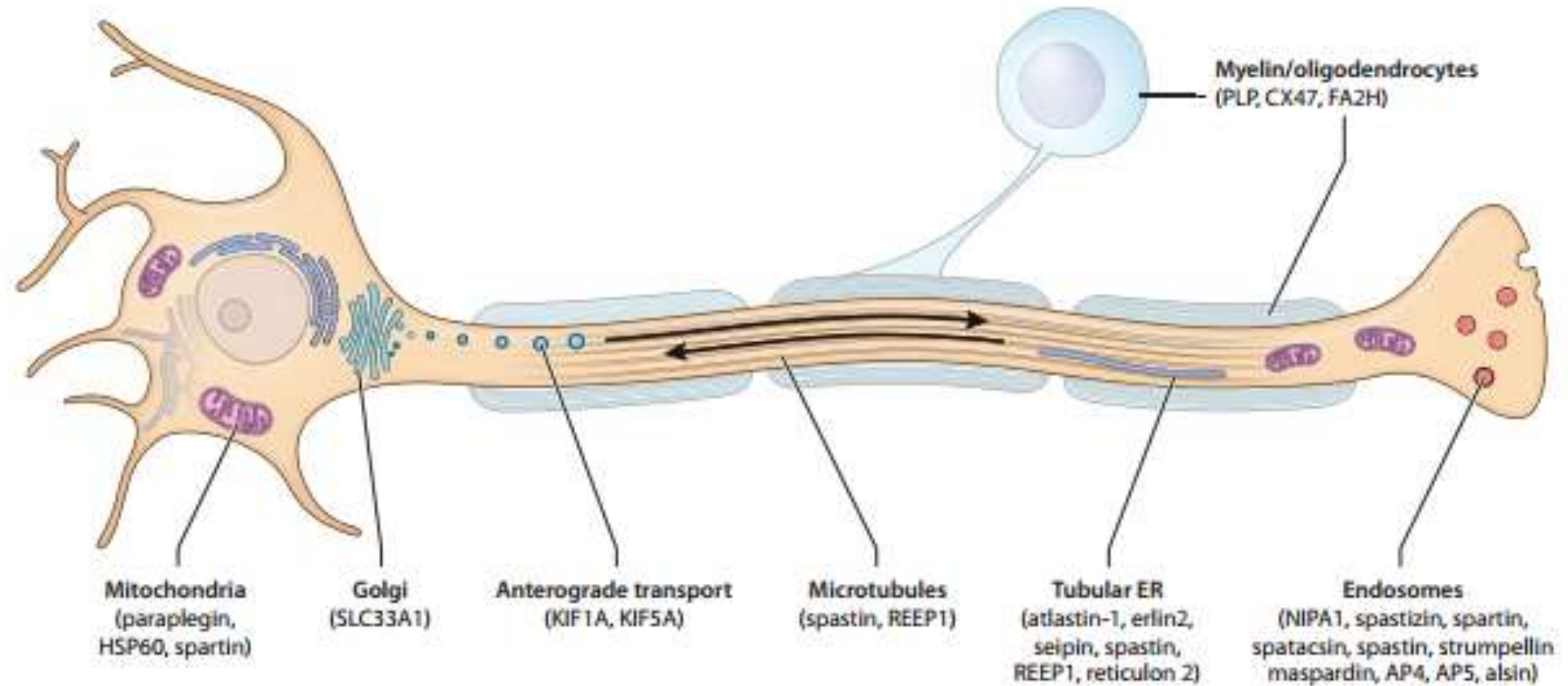
Medical Genetics Unit  
Department of Medical and Surgical Sciences  
University of Bologna



## COMMON FEATURES OF HEREDITARY SPASTIC PARAPLEGIA

- Hereditary neurodegeneration disease
- Clinically and genetically heterogeneous group of disease
- Main neurological signs: spasticity and weakness of the lower limbs
- 80 forms reported (Mendelian inheritance and non)
- Common pathological feature:  
retrograde distal axonopathy of the long descending motor fibers
- Several intracellular pathogenetic mechanisms
- No therapies available, only symptomatic treatments

# Common Pathogenetics Themes in HSPs



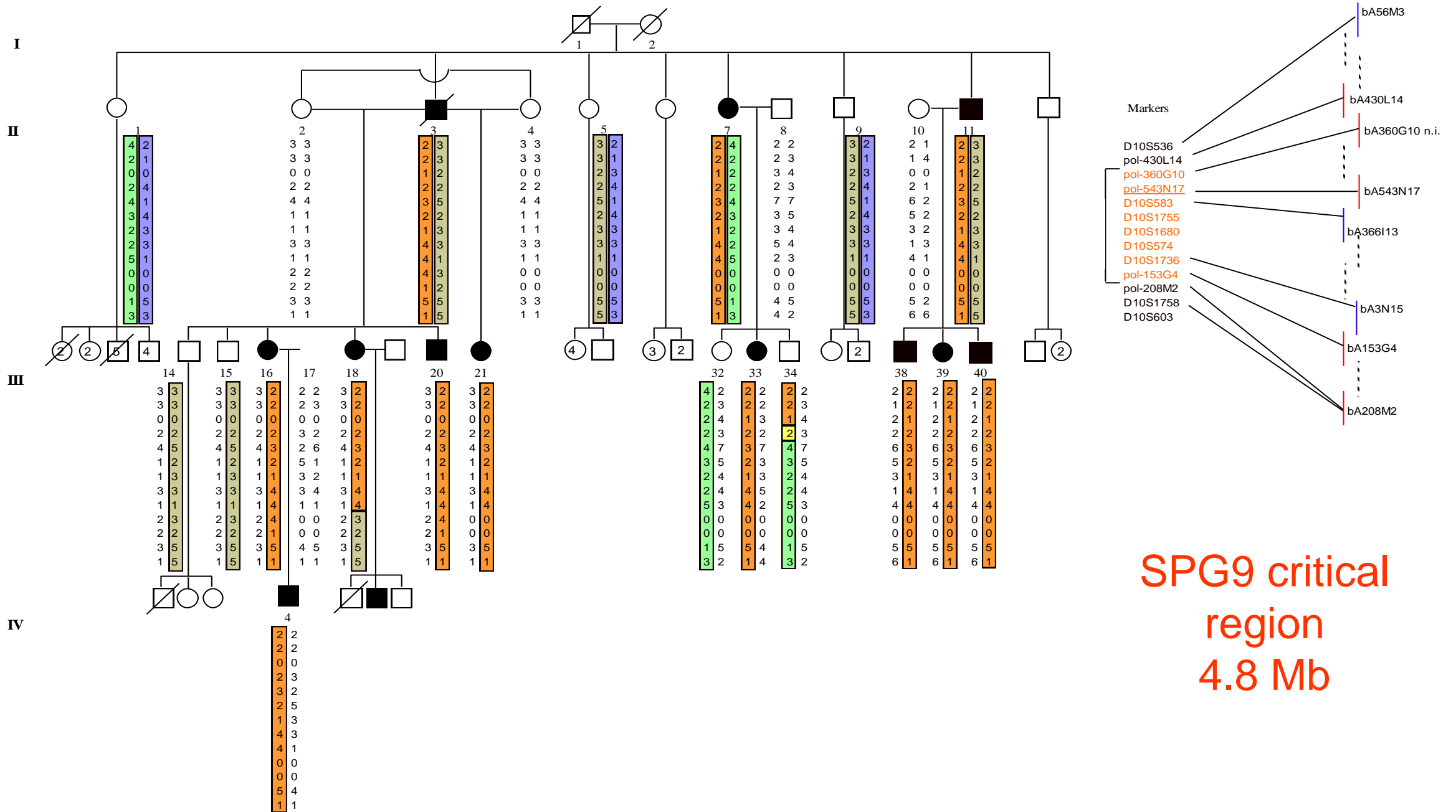
**Genetic Mapping to 10q23.3-q24.2, in a Large Italian Pedigree,  
of a New Syndrome Showing Bilateral Cataracts, Gastroesophageal  
Reflux, and Spastic Paraparesis with Amyotrophy**

Am. J. Hum. Genet. 64:586-593, Seri Marco et al., 1999.

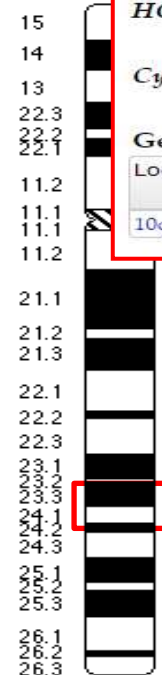
### Main Clinical Features:

- spastic paraplegia with incomplete dominance and/or variable expressivity
- early bilateral cataracts
- persistent vomiting
- autosomal dominant inheritance
- spastic paraplegia arises in the first and third decade of life

# Redefinition of the SPG9 locus



# The SPG9 critical region contains 52 genes



**\*604619**

**LEUCINE-RICH GENE, GLIOM**

*Alternative titles: symbols*  
EPITEMPIN; EPTP

*HGNC Approved Gene Symbol: **LGI1***

*Cytogenetic location: 10q23.33*

*Genomic coordinates (GRCh38): 10:93,757,769-93,798,173 (from NCBI)*

**Gene-Phenotype Relationships**

Location	Phenotype	Phenotype MIM number	Inheritance (in progress)	Phenotype mapping key
10q23.33	Epilepsy, familial temporal lobe, 1	600512	AD	3

**\*138250**

**ALDEHYDE DEHYDROGENASE 18 FAMILY, MEMBER A1; ALDH18A1**

*Alternative titles: symbols*

1-PYRROLINE-5-CARBOXYLATE SYNTHETASE; **PYCS**  
P5CS

GLUTAMATE GAMMA-SEMIALDEHYDE SYNTHETASE; GSAS

*HGNC Approved Gene Symbol: **ALDH18A1***

*Cytogenetic location: 10q24.1*

*Genomic coordinates (GRCh38): 10:95,605,928-95,656,809 (from NCBI)*

- [C10orf129](#)
- [PDLIM1](#)
- [SORBS1](#)
- [LOC439999](#)
- [PYCS](#)**
- [C10orf61](#)
- [ENTPD1](#)
- [FLJ34077](#)
- [LOC387706](#)
- [C10orf130](#)
- [CCNJ](#)
- [ZNF518](#)
- [LOC399804](#)
- [BLNK](#)
- [DNTT](#)
- [TMEM10](#)
- [TLL2](#)
- [SMBP](#)
- [PIK3AP1](#)
- [LOC283340](#)
- [LOC387709](#)
- [MLR2](#)

- [LOC387703](#)
- [FER1L3](#)
- [C10orf3](#)
- [GPR120](#)
- [RBP4](#)
- [PDE6C](#)
- [C10orf4](#)
- [LGI1](#)**
- [TMEM20](#)
- [PSMD4P2](#)
- [PLCE1](#)
- [C10orf117](#)
- [TBC1D12](#)
- [HELLS](#)
- [LOC441573](#)
- [CYP2C18](#)

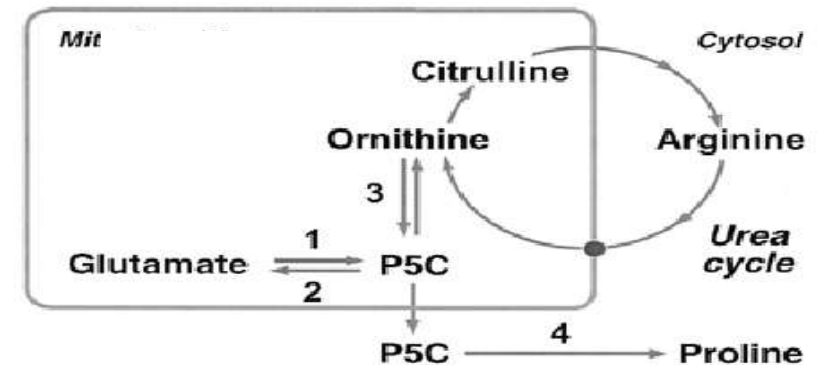
## Hyperammonemia with reduced ornithine, citrulline, arginine and proline: a new inborn error caused by a mutation in the gene encoding $\Delta^1$ -pyrroline-5-carboxylate synthase

Matthias R. Baumgartner<sup>1</sup>, Chien-an A. Hu<sup>1</sup>, Shlomo Almashanu<sup>2</sup>, Gary Steel<sup>2</sup>, Cassandra Obie<sup>2</sup>, Bernard Aral<sup>3</sup>, Daniel Rabier<sup>3</sup>, Pierre Kamoun<sup>3</sup>, Jean-Marie Saudubray<sup>3</sup> and David Valle<sup>1,2,+</sup>

OMIM 138250

Hyperammonemia, ipoornitinemia, ipocitrullinemia, ipoarginemia, ipoprolinemia

Deficit  $\Delta^1$ -pyrroline-5-carbossilato-sintasi





ORIGINAL PAPER

Matthias R. Baumgartner · Daniel Rabier  
Marie-Cécile Nassogne · Jean-Louis Dufier  
Jean-Paul Padovani · Pierre Kamoun · David Valle  
Jean-Marie Saudubray

**$\Delta^1$ -pyrroline-5-carboxylate synthase deficiency: neurodegeneration, cataracts and connective tissue manifestations combined with hyperammonaemia and reduced ornithine, citrulline, arginine and proline**

## SPG9

Persistent vomiting,  
gastroesophageal reflux

Skeletal abnormalities,  
Short stature

Learning disabilities

Bilateral cataracts

Amyotrophy

Motor system disorder  
Spastic Paraparesis

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## Deficit $\Delta^1$ -pyrrolin-5- carbossilato-sintetasi

Chronic vomiting  
(due to gastro-esophageal reflux)

Joint laxity resulting in severe pes planus  
and dislocated hips.  
Slight dysmorphic features  
(short neck long fingers and toes)  
Hyperelasticity of the skin

Mental deterioration and abnormal  
behaviour

Bilateral zonular cataracts

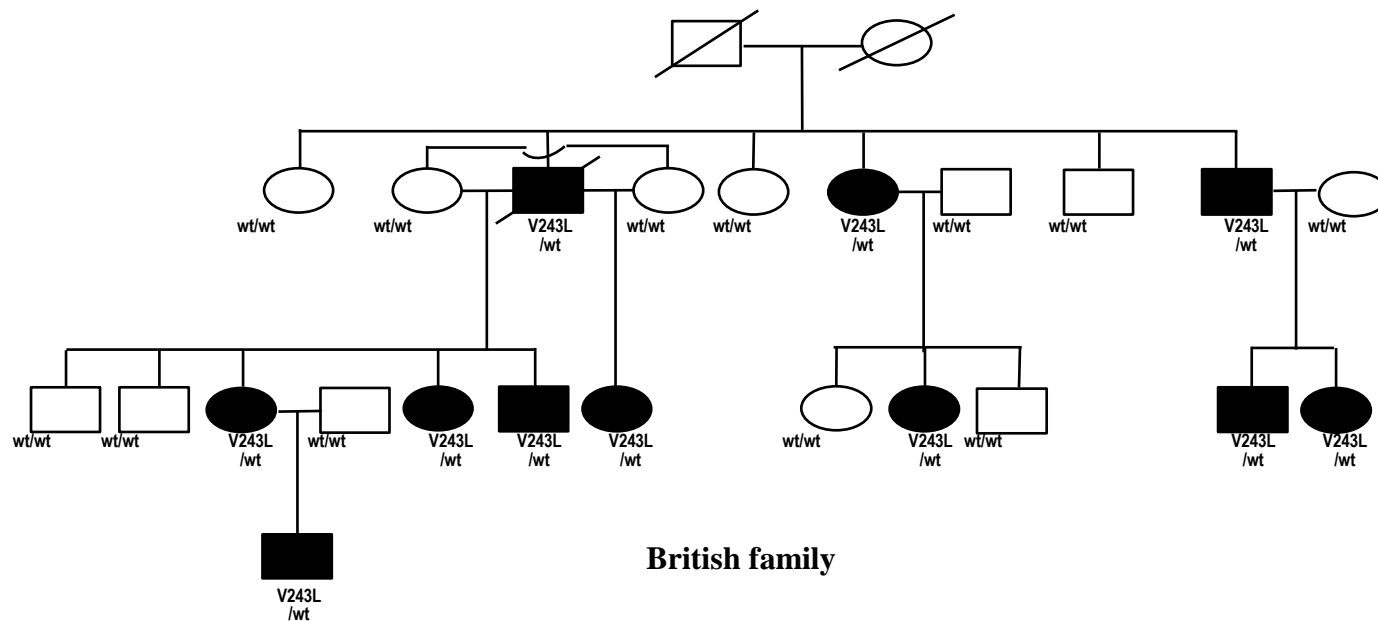
Severe hypotonia, muscular wasting of  
the limbs, dystonia of the hands and feet

Pyramidal syndrome and peripheral,  
predominantly axonal, neuropathy

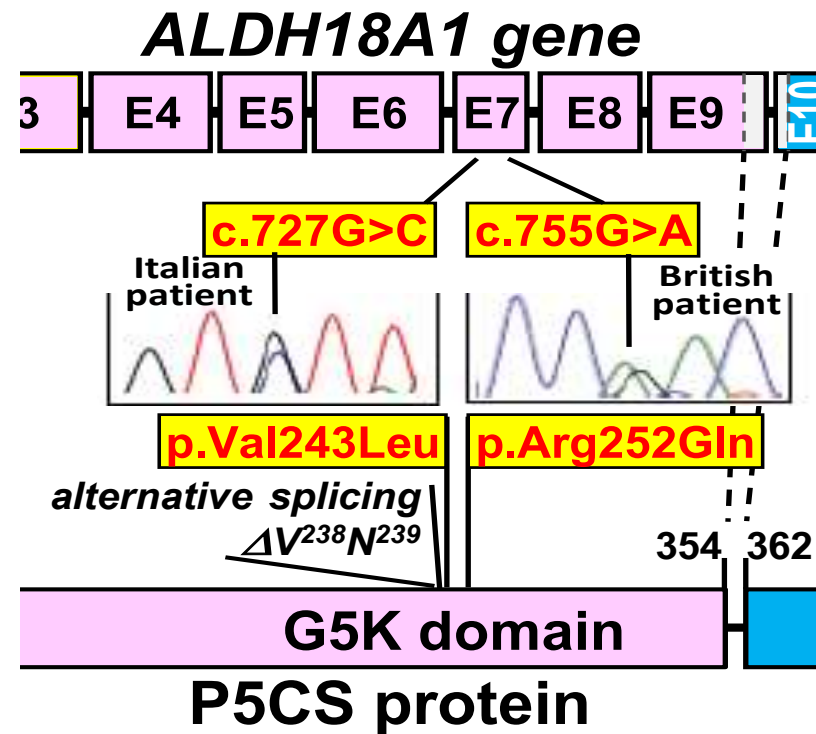
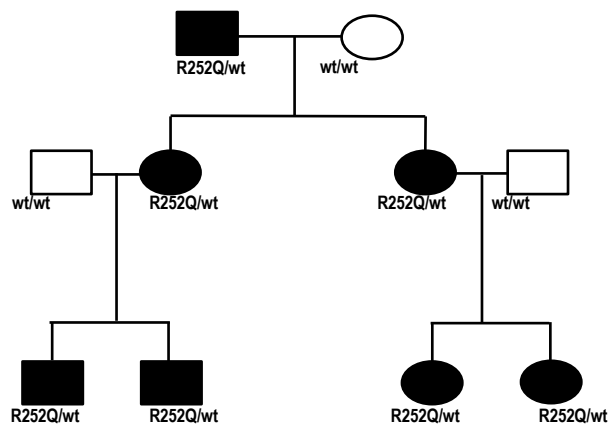
Sanger sequencing and NextGen sequencing confirm the presence of the mutation p.V243L in the Italian family and the mutation p.R252Q in a second English family.

The mutations segregate only in affected patients of the families and are not present in a panel of 466 chromosomes Geographically matched, or in polymorphisms databases (dbSNPs, 1000 Genome Project, Exac).

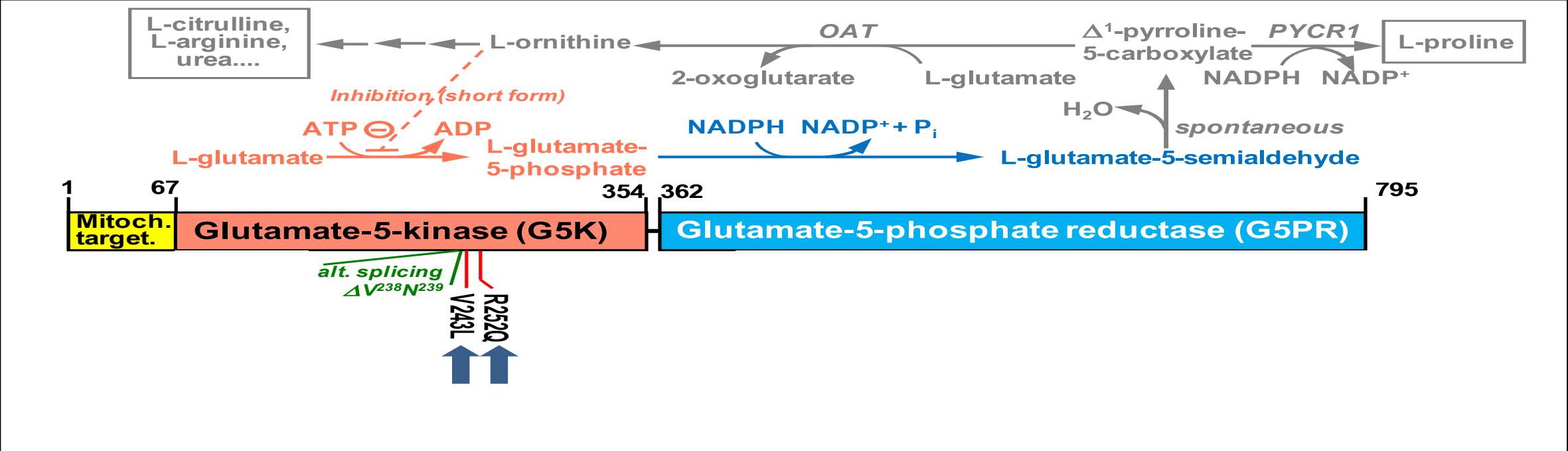
Italian family



British family



P5CS is a byfunctional enzyme catalyzing the first two steps of de novo synthesis of proline, ornithine, citrulline and arginine



# Protein sequence of the mutated region in comparison with the same region in other species

	V243L	R252Q	
	↓	↓	
Homo sapiens	-NVISVKDND	SLAARLAVEMKTDL	261
Mus musculus	-NVISVKDND	SLAARLAVEMKTDL	261
Canis familiaris	-NVISVKDND	SLAARLAVEMKTDL	261
Gallus gallus	-NVISVKDND	SLAARLAVEMKTDL	266
Xenopus laevis	--VISIKDND	SLAARLAVEMKADL	260
Tetraodon nigroviridis	--VISIKDND	SLAARLAVEMKADL	211
Dario rerio	-NVISIKDND	SLAARLAVEMRADL	241
Branchiostoma floridae	-GVISVKDND	SLAARLAAEVQADL	294
Drosophila melanogaster	RRGIPIKDND	SLSAMLAAEVQADL	248
Caenorhabditis elegans	---MHISDND	SLAARLSAEIEAEL	195
Nematostella vectensis	-GVISLKDND	SLAALLAVEIRADL	219
Strongyloectotus purpuratus	-GVISIKDND	SLAARLAEINADL	214
Hydra magnipapillata	-DEIKFGDND	TLGALVANLVEADA	173
Triticum aestivum	----IFWDND	SLAGLLALELKADL	191
Zea mays	----IFWDND	SLAGLLAIELKADL	192
Glycin max	----IFWDND	SLSALLALELKADL	191
Medicago sativa	----IFWDND	SLSALLALELKADL	191
Vigna vinifera	----IFWDND	SLAGLLALQLKADL	193
Vigna unguiculata	----IFWDND	SLAGLLALELKADL	225
Brassica napus	----IFWDND	SLAALLALELKADL	191
Arabidopsis thaliana	----IFWDND	SLAALLSLELKADL	191
Picea sitchensis	----IFWDND	SLAALLALELRADL	188
Physcomitella patens	----IFWDND	SLAALLALELQADL	191
Saccharomyces cerevisiae	-REIKFGDND	TLSAITSAIHHADY	170
Escherichia coli	-AEIKVGDND	NLSALAAIILAGADK	164
Campylobacter jejuni	-EEIVEGDND	SLSAYATHFFDADL	157

## Mutations in SPG9 cause «Loss of Function»

Plasma levels of selected aminoacids in SPG9 patients

**Table 1** Plasma levels of selected amino acids in SPG9 patients

Analyte	Levels in patient and corresponding reference range (in parenthesis) ( $\mu\text{mol/l}$ )					
	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5	Patient 6
Ornithine	72 (41–119)	50 (41–119)	63 <sup>a</sup> (63–133)	63 <sup>a</sup> (63–133)	37 <sup>b</sup> (63–133)	49 <sup>b</sup> (63–133)
Citrulline	17 <sup>a</sup> (17–53)	13 <sup>b</sup> (17–53)	20 (15–57)	21 (15–57)	14 <sup>b</sup> (15–57)	8 <sup>b</sup> (15–57)
Arginine	87 (38–135)	81 (38–135)	101 (36–172)	139 (36–172)	131 (36–172)	80 (36–172)
Proline	186 (117–332)	150 (117–332)	144 (50–350)	215 (50–350)	199 (50–350)	160 (50–350)
Hydroxyproline	5 <sup>b</sup> (16–53)	14 <sup>b</sup> (16–53)	12 (0–50)	12 (0–50)	15 (0–50)	Not determined

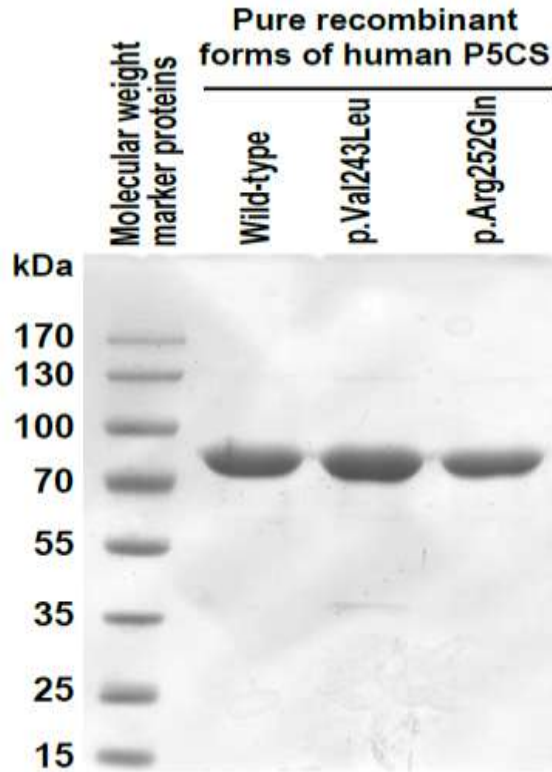
Patients belong to the Italian pedigree and were heterozygotes for the mutation V243L.

<sup>a</sup>Levels are at the lower limit of the reference range.

<sup>b</sup>Levels are below the reference range.

## Mutations in SPG9 cause «Loss of Function»

Mutations in the Human recombinant protein P5CS abolish the activity of the mutated G5K domain



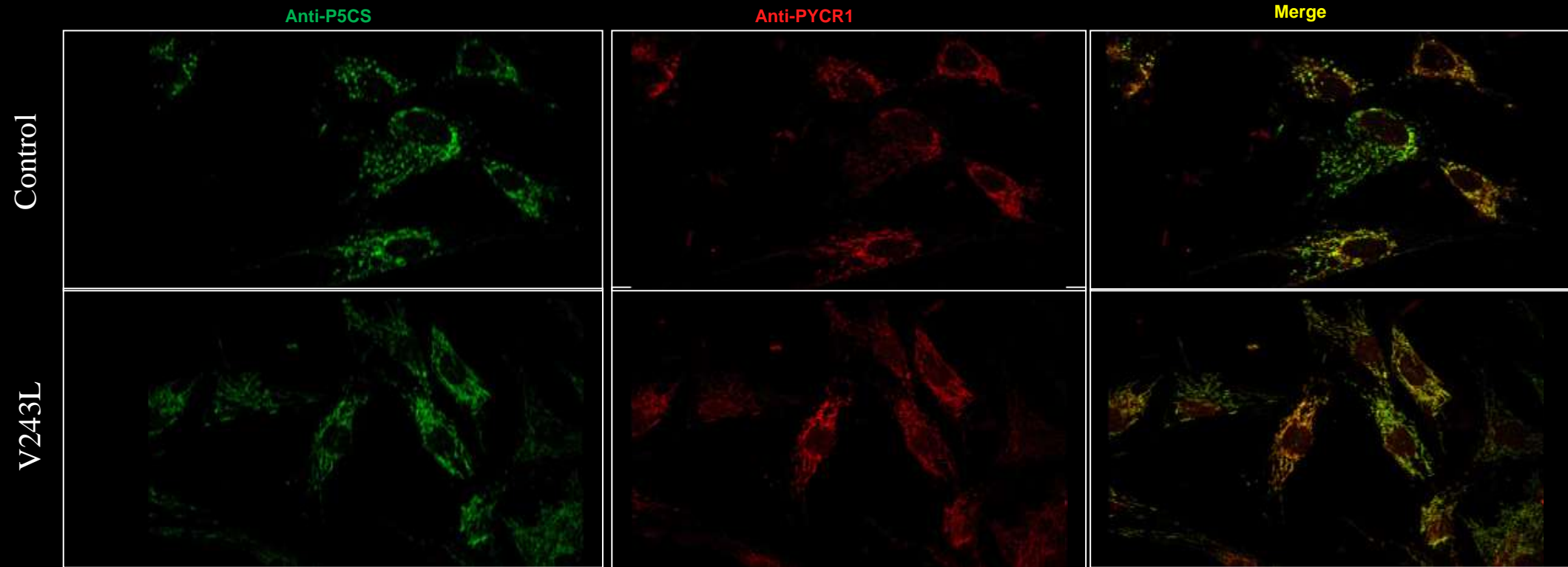
### Enzyme activities of purified recombinant P5CS wild-type and mutant forms

Assay of	Activity U/ mg (% of wild-type)		
	Wild-type	p.Val243Leu	p.Arg252Gln
Total P5CS activity	2.11±0.11 (100)	< 0,07 (< 3.5%)	< 0,07 (< 3.5%)
Partial activities			
G5K	18.3±1.2 (100)	0.018±0.001 (0.1%)	0.027±0.003 (0.15%)
G5PR	2.55±0.07 (100)	1.78±0.01 (70%)	1.94±0.02 (76%)

For details on the assays, see Panza et al. (2016) Brain 139:e3

Immunofluorescence:

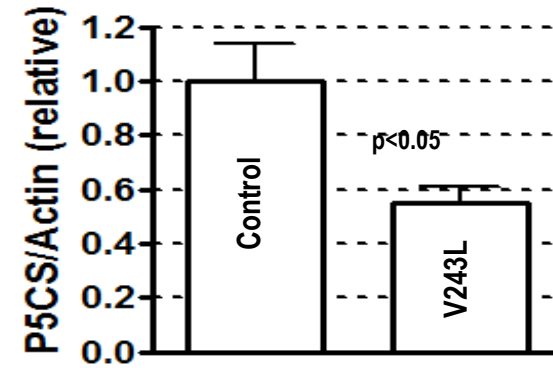
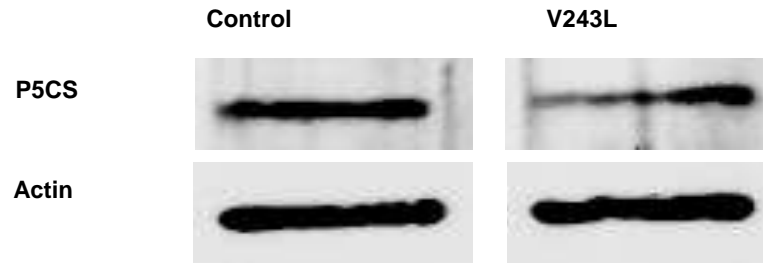
Patient's fibroblasts in a patient of the Italian family bearing the p.V243L mutation



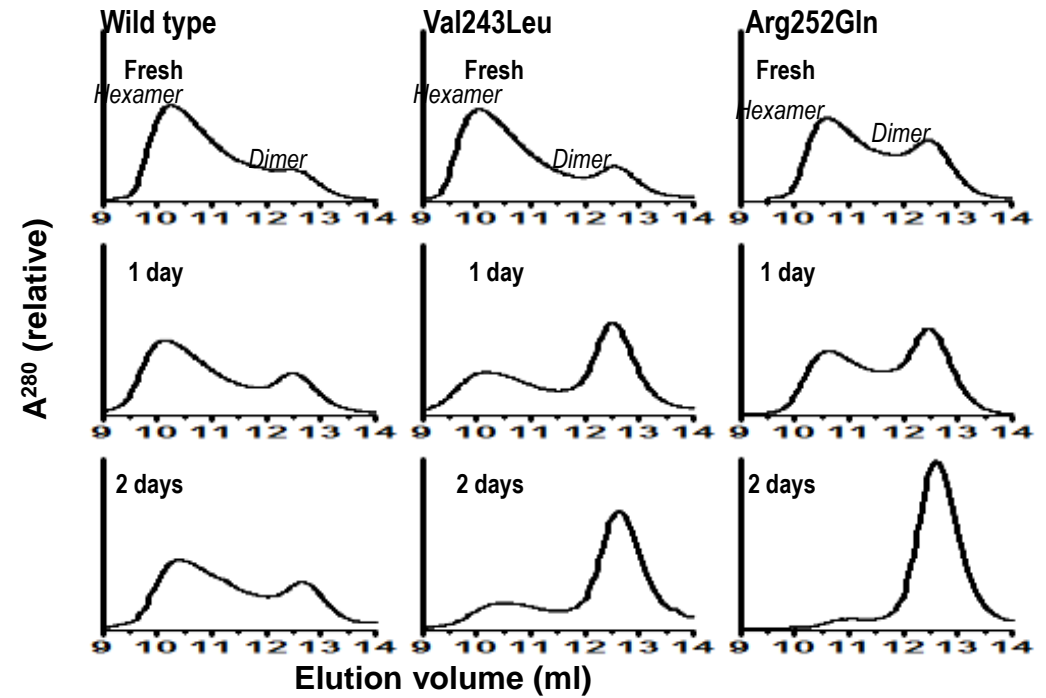
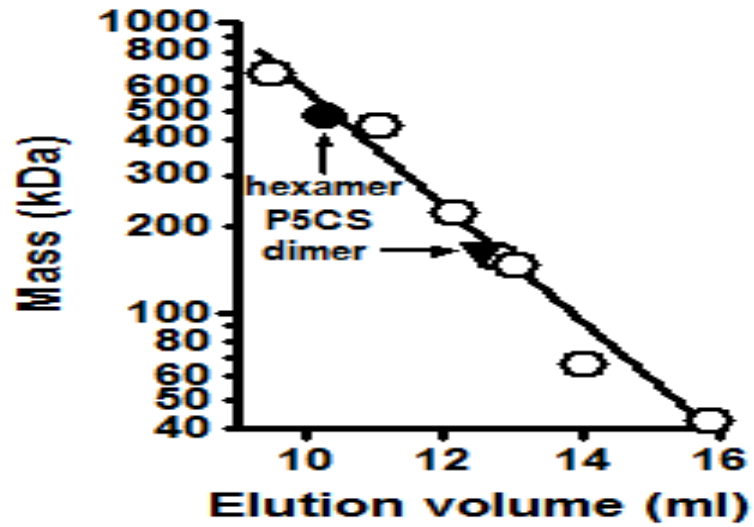
Mutant cDNAs are expressed with a pattern similar to the Wild type and are not associated with abnormalities in the localization nor a premature degradation



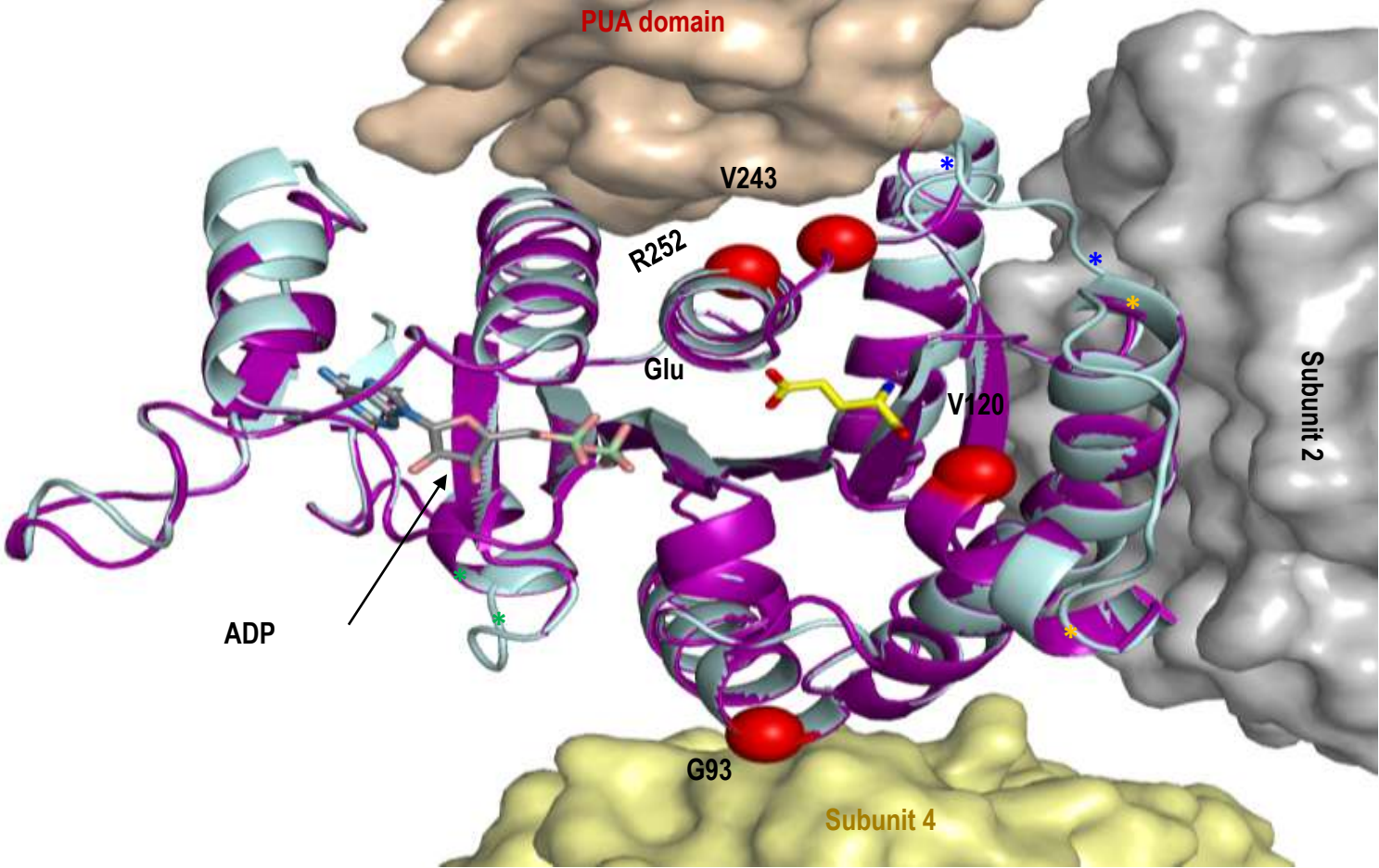
Western blot on patient's fibroblasts and controls show a roughly 45% reduction of P5CS in p.V243L cells



Why a dominant inheritance for «Loss of Function» mutations?



Why dominant and recessive mutations exist?



# Recessive and dominant mutations in *ALDH18A1* have been identified in forms of HSP and of cutis laxa

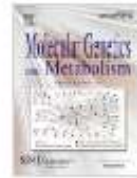
ARCL3A OMIM 219150



ELSEVIER

Molecular Genetics and Metabolism

Volume 112, Issue 4, August 2014, Pages 310–316



## Severe congenital cutis laxa with cardiovascular manifestations due to homozygous deletions in *ALDH18A1*

Björn Fischer<sup>a, b</sup>, Bert Callewaert<sup>c</sup>, Phillippe Schröter<sup>a</sup>, Paul J. Coucke<sup>c</sup>, Claire Schlack<sup>a</sup>, Claus-Eric Ott<sup>a, b</sup>, Manrico Morroni<sup>d</sup>, Wolfgang Homann<sup>e</sup>, Stefan Mundlos<sup>a, b, f</sup>, Eva Morava<sup>g</sup>, Anna Ficcadenti<sup>h</sup>, Uwe Kornak<sup>a, b, f</sup>.  

ADCL3 OMIM 616603

AJHG

Volume 97, Issue 3, 3 September 2015, Pages 483–492



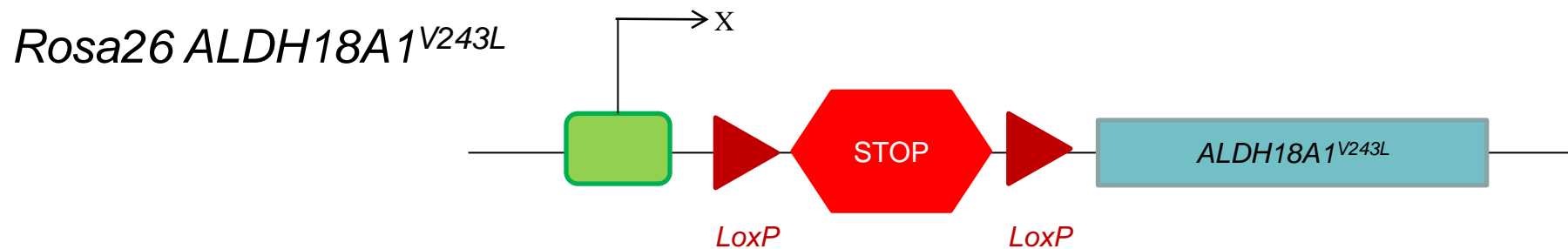
Report

## Recurrent De Novo Mutations Affecting Residue Arg138 of Pyrroline-5-Carboxylate Synthase Cause a Progeroid Form of Autosomal-Dominant Cutis Laxa

Björn Fischer-Zirnsak<sup>1, 2, 25</sup>, Nathalie Escande-Beillard<sup>3, 25</sup>, Jaya Ganesh<sup>4</sup>, Yu Xuan Tan<sup>3</sup>, Mohammed Al Bughaili<sup>1</sup>, Angela E. Lin<sup>5</sup>, Inderneel Sahai<sup>5</sup>, Paulina Bahena<sup>6</sup>, Sara L. Reichert<sup>4</sup>, Abigail Loh<sup>7</sup>, Graham D. Wright<sup>3</sup>, Jaron Liu<sup>3</sup>, Elisa Rahikkala<sup>8</sup>, Eniko K. Pivnick<sup>9</sup>, Asim F. Choudhri<sup>10, 11, 12, 13</sup>, Ulrike Krüger<sup>1</sup>, Tomasz Zemojtel<sup>1, 14</sup>, Conny van Ravenswaaij-Arts<sup>15</sup>, Roya Mostafavi<sup>9</sup>, Irene Stolte-Dijkstra<sup>15</sup>, Sofie Symoens<sup>16</sup>, Leila Pajunen<sup>8</sup>, Lihadh Al-Gazali<sup>17</sup>, David Meierhofer<sup>18</sup>, Peter N. Robinson<sup>1, 2, 19</sup>,

# Towards the generation of a mouse model for «*ALDH18A1*-related disease»

- To investigate the pathogenetic mechanism of this disease
- To test new therapies



Generated in Mario Capecchi Laboratory

## Summary

SPG9 is due to the non-synonymous single nucleotide changes c.727G>C or c.755G>A in exon 7 of the ALDH18A1 gene that affect proximate amino acids of the G5K domain (p.Val243Leu and p.Arg252Gln, respectively).

These mutations do not prevent production nor cause cellular mislocalization of P5CS.

They are loss-of function mutations as evidenced by plasma amino acid analysis and by enzyme activity studies in recombinantly produced human P5CS.

They selectively inactivate the domain where they map (G5K).

P5CS is a high oligomer (possibly an hexameric trimer of dimers).

SPG9 mutations disturb the architecture of the P5CS oligomer, making it prone to dissociate to dimers.

In silico structural analysis suggests that P5CS mutations can be dominant or recessive depending on whether they affect or not residues involved in intersubunit or interdomain interactions, disturbing or not disturbing the architecture of the oligomer, thus supporting a dominant negative disease-causing mechanism

Recessive and dominant mutations have been identified in HSP (SPG9A MIM601162, SPG9B MIM616586) and in forms of cutis laxa (ADCL3 MIM616603, ARCL3A MIM219150)

The generation of a mouse model will be essential to dissect the pathogenetic mechanisms of SPG9 and to test new therapies

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CIBERER-ISCIII, Valencia-Spain

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